



An Introduction to Laboratory for TB Nurses

Lisa Armitige, MD, PhD

Assistant Medical Director
Heartland National TB Center

Associate Professor of Medicine/Pediatrics
UT Health Science Center at Tyler

TB Diagnostic Methods


- **IGRA (Interferon Gamma Release Assay)**
- **AFB Smear**
- **Nucleic Acid Amplification**
- **AFB Culture**
- Clinical Presentation
- TST (Tuberculin Skin Test)
- X-ray

IGRAs

- Which tube do I use?
 - T-spot (heparin – green top tube)
 - QFT – tubes that come with the kit or a heparin – green top (if your lab will let you.....)
- Which lab do I send it to? How do I get it there?
- How quickly do I need to have it to the lab?
- How do I store the tube until transport?



QuantiFERON[®]-TB Gold Plus



Mitogen – Positive Control
Low response may indicate inability to generate IFN- γ
Nil – Negative Control
Adjusts for background IFN- γ
TB1 – Primarily detects CD4 T cell response
TB2 – Optimized for detection of CD4 and CD8 T cell responses



- Essentially 2 tests in one blood draw
- TB1 and TB2 should be close in value

Sputum specimens

- Who should I collect sputum from
 - Patients with respiratory symptoms
 - Patients with an abnormal CXR



Bacteriologic and histologic Examinations

Especially when lung or larynx is site of disease:

- 3 sputum specimens for AFB smear and culture
- Collected 8-24 hours apart with at least 1 early morning specimen



***Specimens should be obtained
in an isolated, well-ventilated
area or sputum collection booth***

Bacteriologic and histologic Examinations

- Sputum collection should be directly supervised
- For patients unable to cough up sputum, deep coughing may be induced

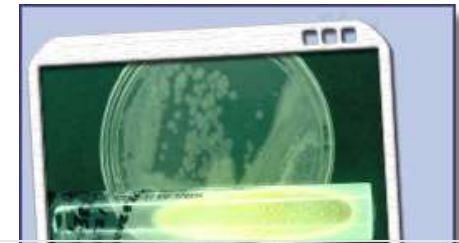


Bacteriologic and histologic Examinations

Extrapulmonary Specimens

- Urine
- Cerebrospinal fluid *
- Pleural fluid *
- Pus
- Biopsy specimens

*recovery poor



**Do NOT collect
specimens in Formalin**



Laboratory Examination

AFB Smear

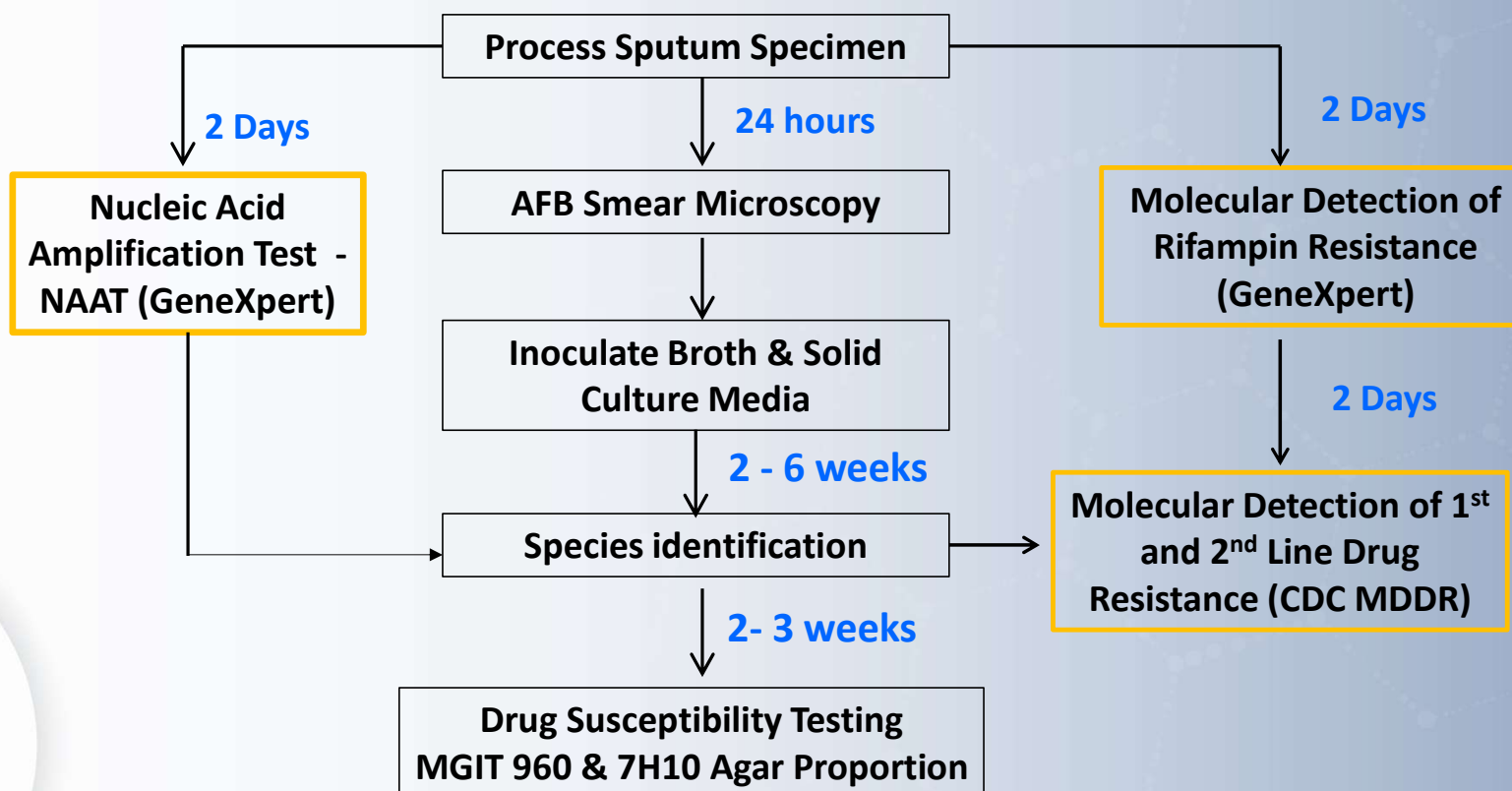
- First clue
- Presumptive diagnosis only
- Fluorochrome staining preferred method
- Results available in 24 hours
- Many patients have negative AFB smears



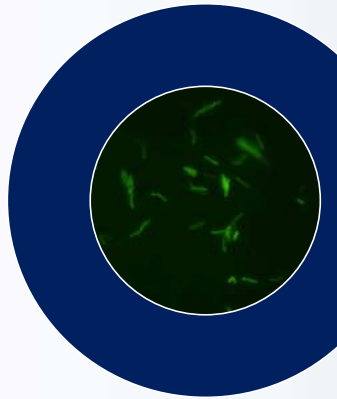
Specimen Quality

- Accurate laboratory results are directly proportional to the quality of the specimen
- Sputum
 - *Recently* discharged material from the bronchial tree, with minimal amounts of upper respiratory tract secretions
 - Well coached patient, collect at least 3 ml
 - Label tube, form, and indicate test:
 - Initial Dx: Smear, NAAT, & Culture
 - Follow-up: Smear and Culture
 - Release from respiratory isolation?
 - Order Smear only
- Transport to lab cool and quickly

TB Laboratory Testing Algorithm



AFB Smear



one
microscopic
field

CAP	ATS	Interpretation	AFB/ml sputum	Infectiousness of patient
negative	negative	negative	<5,000	probably not infectious
1 or 2 per smear	1 or 2 per smear	weakly positive	~5,000	probably infectious
<1 per field	1+	moderately positive	~10,000	probably infectious
1-10 per field	2+	moderately positive	~100,000	probably infectious
	3+	strongly positive	~1,000,000	probably very infectious
>10 per field	4+	strongly positive	>1,000,000	probably very infectious

Nucleic Acid Amplification Tests (NAAT)

- Tiny amounts of DNA/RNA are amplified (copied) until there is enough for easy detection
- DNA/RNA is examined
 - Identification
 - Detection of Drug Resistance
- Test turnaround time measured in hours

Nucleic Acid Amplification Test (NAAT)

- Detects *M. tuberculosis* complex nucleic acids; **does not distinguish between live and dead bacilli**
 - For initial diagnosis only
 - Not suitable for follow-up specimen or monitoring; cured patients may be NAAT + for years!
- Sensitivity compared to TB culture
 - >95% for AFB smear-positive
 - Only 55-75% for AFB smear-negative
- Does not replace culture for bacteriological Dx

Laboratory Examination

Cultures

Used to confirm diagnosis

- Perform on ALL specimens regardless of AFB smear results
- Results available in 10 to 14 days (on liquid media, e.g. BACTEC)

TB may be diagnosed on the basis of signs and symptoms in the absence of a + culture



AFB Culture

- More sensitive than smear
 - 5,000 to 10,000 AFB/ml for smear
 - ~10 viable AFB/ml for culture
- Positive for only ~85% of Pulmonary TB
 - Requires a quality specimen
 - May be invalid due to contamination
- Used to monitor patient response to treatment (like smear)
- Required for drug susceptibilities & genotype
- Lengthy
 - 1-6 weeks by liquid media
 - 2-8 weeks by solid media

Drug Susceptibility Testing (DST) of *M. tuberculosis* complex

Current Recommendations

- Initial isolate should be tested against first-line drugs (FLD)
 - Isoniazid, Rifampin, Ethambutol, Pyrazinamide
 - Repeat test if patient cult+ after 3 mo. Rx
- For isolates resistant to Rifampin or to any 2 FLDs: **test second-line drug panel**
 - Minimum: Fluoroquinolone, Ethionamide, & Injectable (Amikacin, Capreomycin, Kanamycin)

Molecular Detection of Drug Resistance

- Examining DNA of specific genes for mutations known to be associated with phenotypic resistance
- Rapid - analysis takes less than 1 day
- Can be done on culture isolates or directly on NAAT+ specimens



CDC Molecular Detection of Drug Resistance (MDDR)

• Test Indications

- Known/suspect DR case or contact to DR case
- Previous TB Treatment
- Patient from area with high rate of DR TB
- Large public health impact
- Mixed or nonviable culture



CDC Molecular Detection of Drug Resistance (MDDR)

- Provides 2-3 day DNA sequence analysis for drug resistance prediction
 - 7 classes of anti-TB drugs sequenced
- MDDR complements conventional DST
 - Used alone, MDDR and conventional DST are imperfect
 - Used together, accuracy of drug resistance or susceptibility detection can be improved.
- Conventional DST results are still needed to confirm susceptibility to individual drugs.

Summary

- Make friends with the laboratory that processes your specimens. Often if you can tell them what you are trying to do, they will help you get there
- Like most things we do, quality matters. That goes for the specimens that are sent to the laboratory
- Molecular tests are one of the biggest jumps forward in information informing patient decisions



Big Thanks to Denise Dunbar from the
Texas State Lab for use of her slides

Questions?

Lisa.Armitige@dshs.texas.gov

1-800-TEX-LUNG

