



**“I would like to order an MDDR with a  
side of MIC and WGS. What is your  
traditional DST today?”**

Benjamin Alpers & Jan Owen  
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New Directions in TB  
April 1 – 2, 2024  
Houston, Texas

**Benjamin Alpers** has the following disclosures to make:

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- No conflict of interests
- No relevant financial relationships with any commercial companies pertaining to this educational activity



**Jan Owen** has the following disclosures to make:

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- No conflict of interests
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**TEXAS**  
Health and Human  
Services

**Texas Department of State  
Health Services**

**“I would like to order an MDDR with a side of MIC and WGS. What is your traditional DST today?”**

or

Making Sense of the TB Diagnostic Menu

**Benjamin Alpers, Applications Scientist/TB Reference Team Lead**

**Jan Owen, Mycobacteriology/Myiology Branch Manager**

**DSHS Austin Laboratory**

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# AFB Smear

- Used in diagnosis for 100+ years
- Rapid and universally available
  - We strive for 24hr TAT
- Identifies priority specimens for NAAT
  - 5,000 to 10,000 AFB/ml must be present in specimen to be detected in smear
- Not sensitive
  - Misses ~50% of TB
- Not specific
  - Positive smear may be NTM
- Can detect dead bacilli



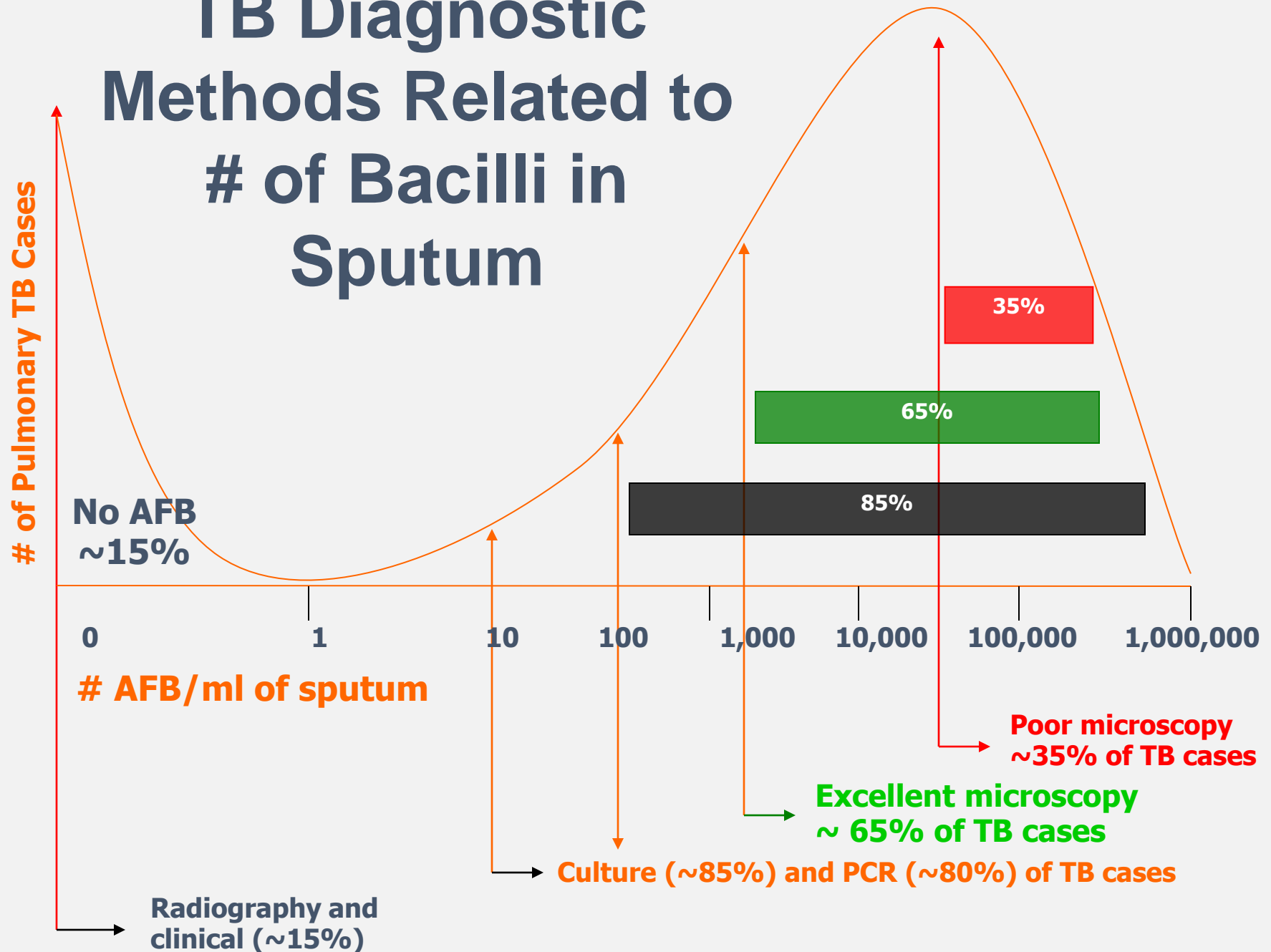
# AFB Culture

- More sensitive than smear
  - ~10 viable AFB/ml for culture
- Positive for ~85% of Pulmonary TB
  - Requires a quality specimen
  - May be invalid due to contamination
- Used to monitor patient response to treatment (like smear)
- Required for conventional drug susceptibilities & genotype
- Lengthy
  - 1-6 weeks by liquid media
  - 2-8 weeks by solid media





# TB Diagnostic Methods Related to # of Bacilli in Sputum



Texas Department of State Health Services

# Nucleic Acid Amplification Tests (NAAT)

- Real time reverse transcription polymerase chain reaction (qRT-PCR or qPCR)
- Tiny amounts of DNA/RNA amplified (copied) until a significant signal compared to the background
- GeneXpert examines DNA for:
  - Identification
  - Detection of Rifampin Resistance
- Test turnaround time measured in hours

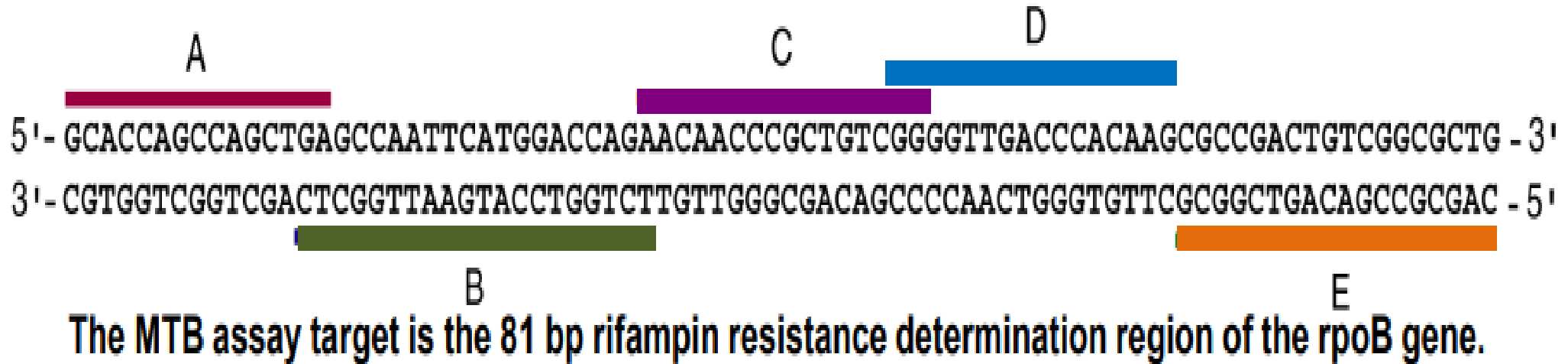


# Nucleic Acid Amplification Tests (NAAT) cont.

- Detects *M. tuberculosis* complex nucleic acids
  - For initial diagnostic specimens only
  - Does not distinguish between live and dead bacilli
  - Not suitable for follow-up specimen or monitoring; cured patients may be NAAT + for years!
- Xpert sensitivity compared to TB culture
  - >95% for AFB smear-positive
  - Only 55-75% for AFB smear-negative



# Cepheid GeneXpert® Target Region



Approx. 10% of rifampin resistant predictions are false  
(ex. Phe433Phe silent mutation)

**GX rifampin resistant results must be confirmed by  
MDDR testing at the CDC.**



# Methods for Culture ID

- Lab-developed, species-specific PCR tests
- Matrix-assisted laser desorption/ionization time of flight (MALDI-TOF or MALDI)
- High-performance liquid chromatography (HPLC)



# Pro and Cons of ID Methods

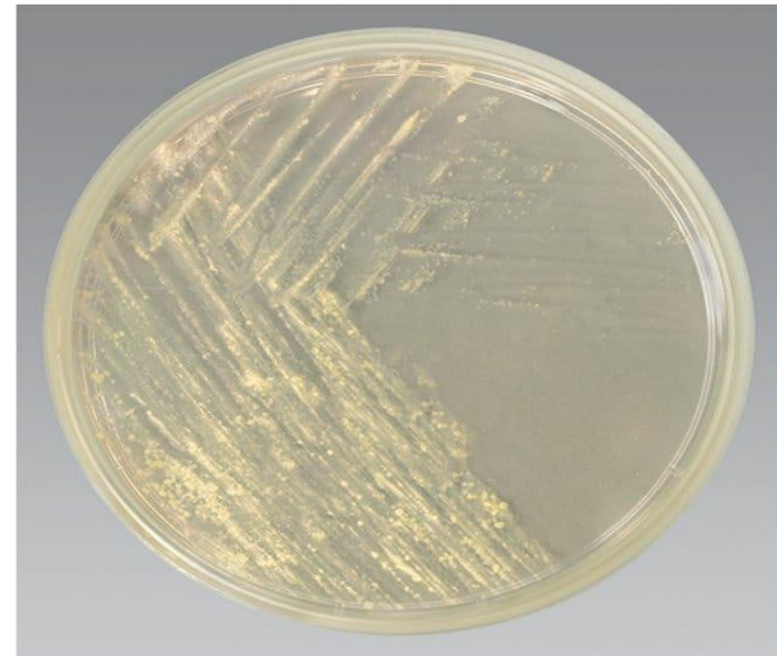
- PCR highly sensitive but costly
  - Can only ID organism for which the assay was developed
- MALDI very specific
  - Can identify extremely wide range of organisms
  - Relatively poor sensitivity; requires substantial growth
- HPLC sensitive, inexpensive, and non-labor intensive
  - Chromatographs require highly trained interpretation
  - Often cannot distinguish organisms within complex or group



# M. tb cx. Speciation through Real-Time Polymerase Chain Reaction (PCR)

Can differentiate between species within the complex using 5 sets of probes and primers targeting known regions of difference (RD)

- *M. tuberculosis*
  - *M. bovis*
  - *M. bovis* BCG
- } inherently PZA resistant
- and
- *M. africanum*
  - *M. microti*
  - *M. canettii*
  - *M. caprae*
- ...and others



# Conventional, Growth-based Drug Susceptibility Testing (DST)

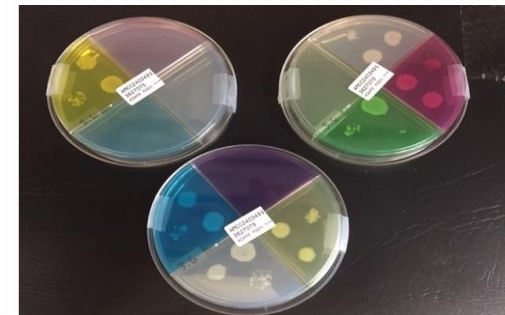
## MGIT 960 method

- Automated
- Widely available
- Poor accuracy
- Easily contaminated



## Agar Proportion method

- Highly sensitive
- Inexpensive production
- Interpretation requires extensive experience



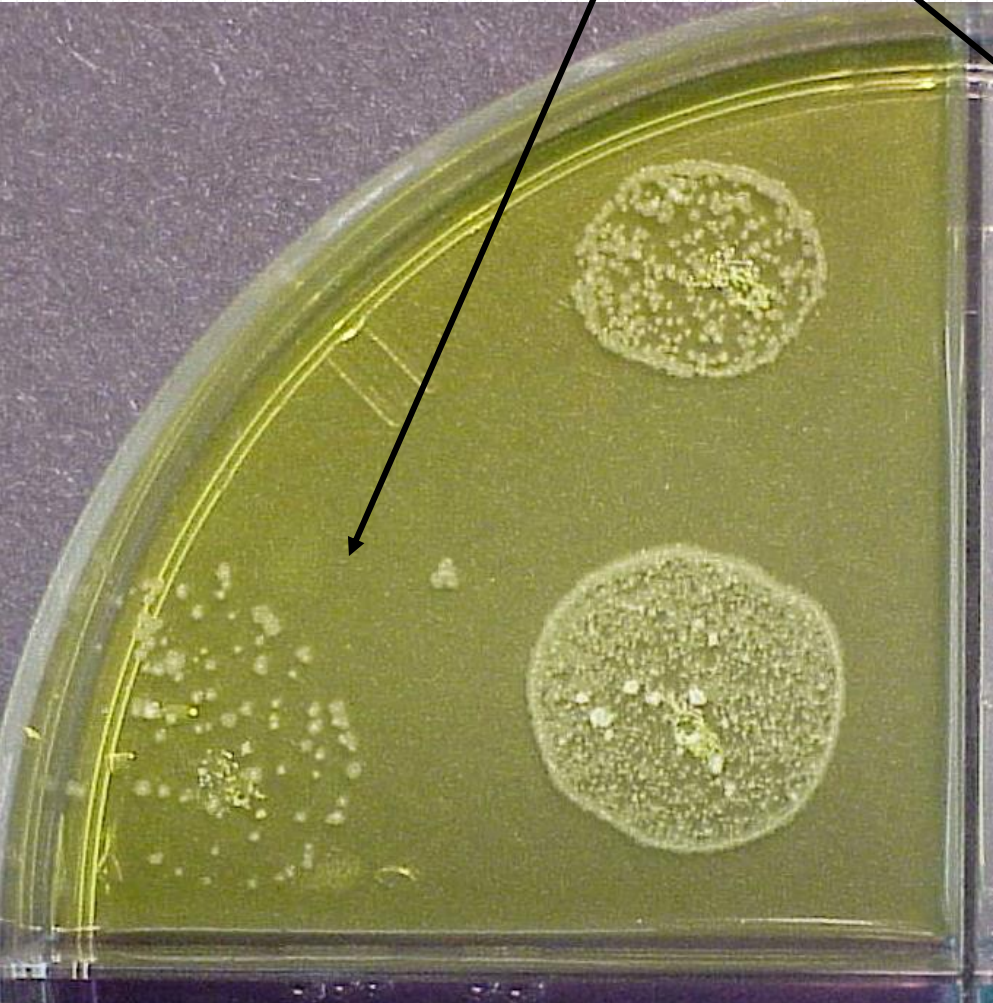


# Agar Proportion (AP) DST Method Principle

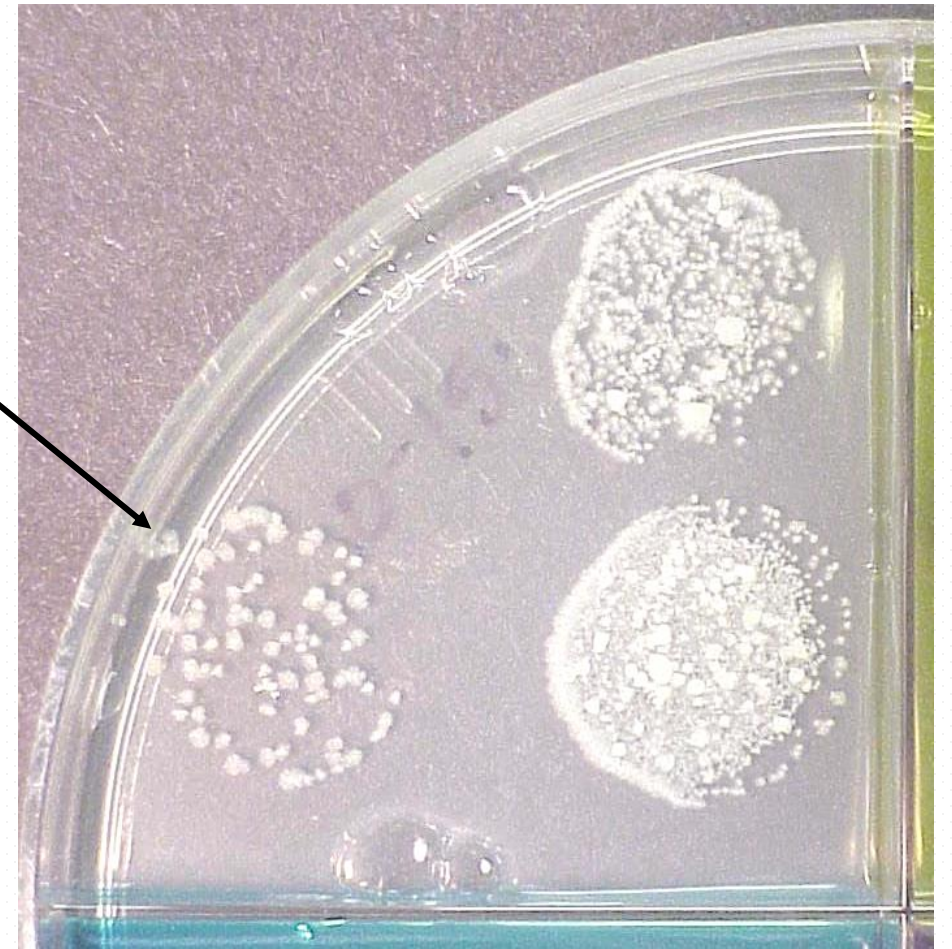
- Standardized suspension inoculated to quadrant plates
  - Drug-containing Middlebrook 7H10 agar
  - Drug-free control
- Drug no longer counted on as being effective if:
  - Growth of on drug quadrant is 1% or greater than the growth on the control



**100cfu/100cfu = 100% Resistance**



Isoniazid, 1.0 mcg/ml



Drug-Free Control

# Additional Considerations

- CDC recommends RMP DST results be reported within 17 days after *M. tuberculosis* culture identification
- Most U.S. laboratories use a rapid commercial system for DST (MGIT 960)
- Commercial DST methods miss some clinically significant RMP resistance that can be detected by agar proportion
- Agar Proportion (AP) is the “gold standard” method for conventional DST...however AP is not a rapid method; conventional AP method takes 21 days (3 weeks) for full results
- DSHS regularly reports INH & RMP susceptibility within 17 days of culture identification
- PZA testing can only be performed by MGIT 960 method



# Minimum Inhibitory Concentration (MIC)

- Lowest concentration of a drug which prevents detectable in vitro growth when tested in a series of concentrations
- Only available at specialized laboratories such as CAHD and Wadsworth Center NYHD
- Performed only by special request on select antibiotics
- Tested by MGIT, AP, or Broth Micro-dilution (BMD)



# Types of Molecular Sequencing

Required DNA Volume



Pyrosequencing (PSQ)

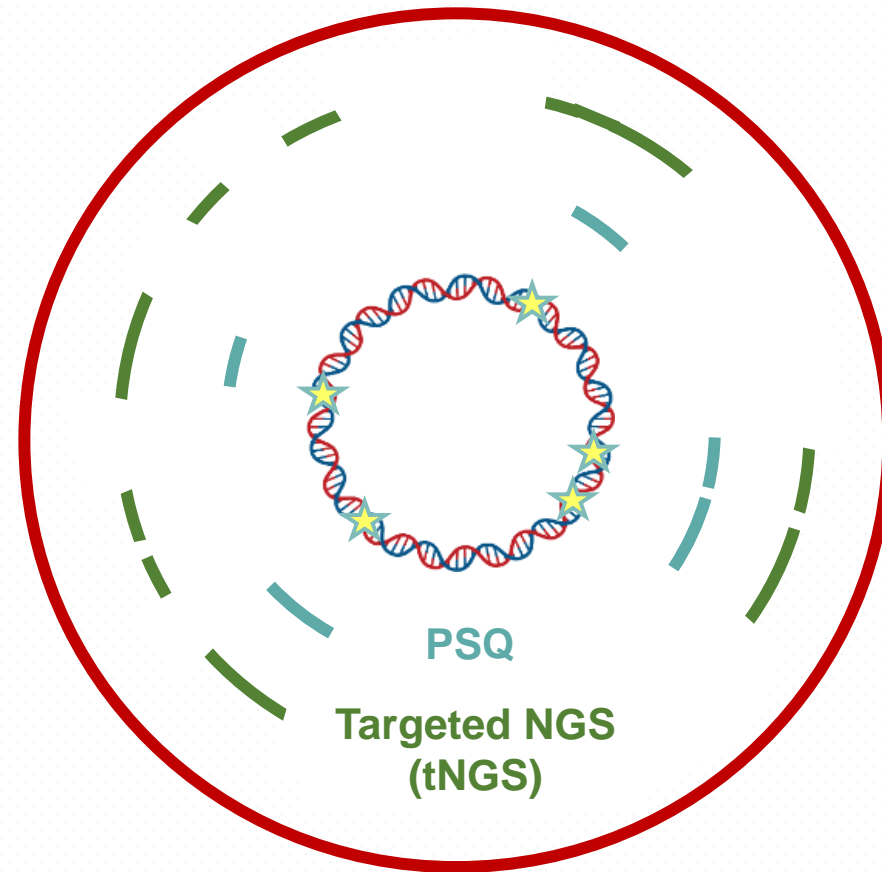
Sanger Sequencing

Targeted Next Generation Sequencing  
(tNGS)

Whole Genome Sequencing (WGS)



# Molecular methods of resistance detection: difference in purview



## Whole Genome Sequencing (WGS)

Adapted from Whole Genome Sequencing for TB Drug Resistance at the TB DST Reference Center, 2024, APHL



Texas Department of State  
Health Services

# Pyrosequencing

- Highly sensitive
- Amplifies short sequences of base pairs
- Quick turnaround (3-5 days)
- Can be performed on PCR+ sediment with very low smears
- CAHD uses if WGS unobtainable (sediment or mixed culture)
- Wadsworth tests new patient PCR+ and to confirm primary or fluroquinolone resistance



# Sanger Sequencing

- Requires ~10x more DNA than pyrosequencing
- Amplifies longer sequences
- Turnaround (5-7 days)
- Can be performed on PCR+ sediment with low smear count
- CDC sequences *rpoB* if tNGS fails or delayed
- Wadsworth sequences *pncA* (PZA) by this method





# tNGS Sequencing

- Requires slightly more DNA than Sanger
- Amplifies longer and more sequences
- Turnaround 7-10 days
- Can be performed on PCR+ sediment with smear positivity of at least 1-10/field
- Can detect subpopulations as low as 10%
- Primary CDC sequencing scheme
- CAHD plans to bring online in near future
- FLHD currently performs



# Genetic Loci Sequenced through tNGS MDDR (CDC)

## Genetic Locus

RRDR within the *rpoB* gene with  
the addition of two codons  
outside of the RRDR

*inhA, katG, fabG1*

*embB*

*pncA*

*gyrA, gyrB*

*rrs*

*eis*

## Associated Drug

Rifampin (RMP)

Isoniazid (INH)

Ethambutol (EMB)

Pyrazinamide (PZA)

Fluoroquinolones

Amikacin Capreomycin

Kanamycin

Kanamycin



# Additional Genetic Loci Sequenced through tNGS

## Genetic Locus

## Associated Drug

*atpE*

*rv0678*

*pepQ*

Bedaquiline

*rv0678*

*pepQ*

Clofazimine

*rplC*

*rrl* (partial)

Linezolid



# WGS Sequencing

- Requires culture (sediment cannot amplify)
- Sequences practically entire genome
- Turnaround 12-16 days
- All major laboratories perform (CA, NY, FL, TX, CDC)
- Can be used for both DST and TB species ID
- Extremely helpful for epidemiological investigation



# TB WGS-DST targets (CAHD)

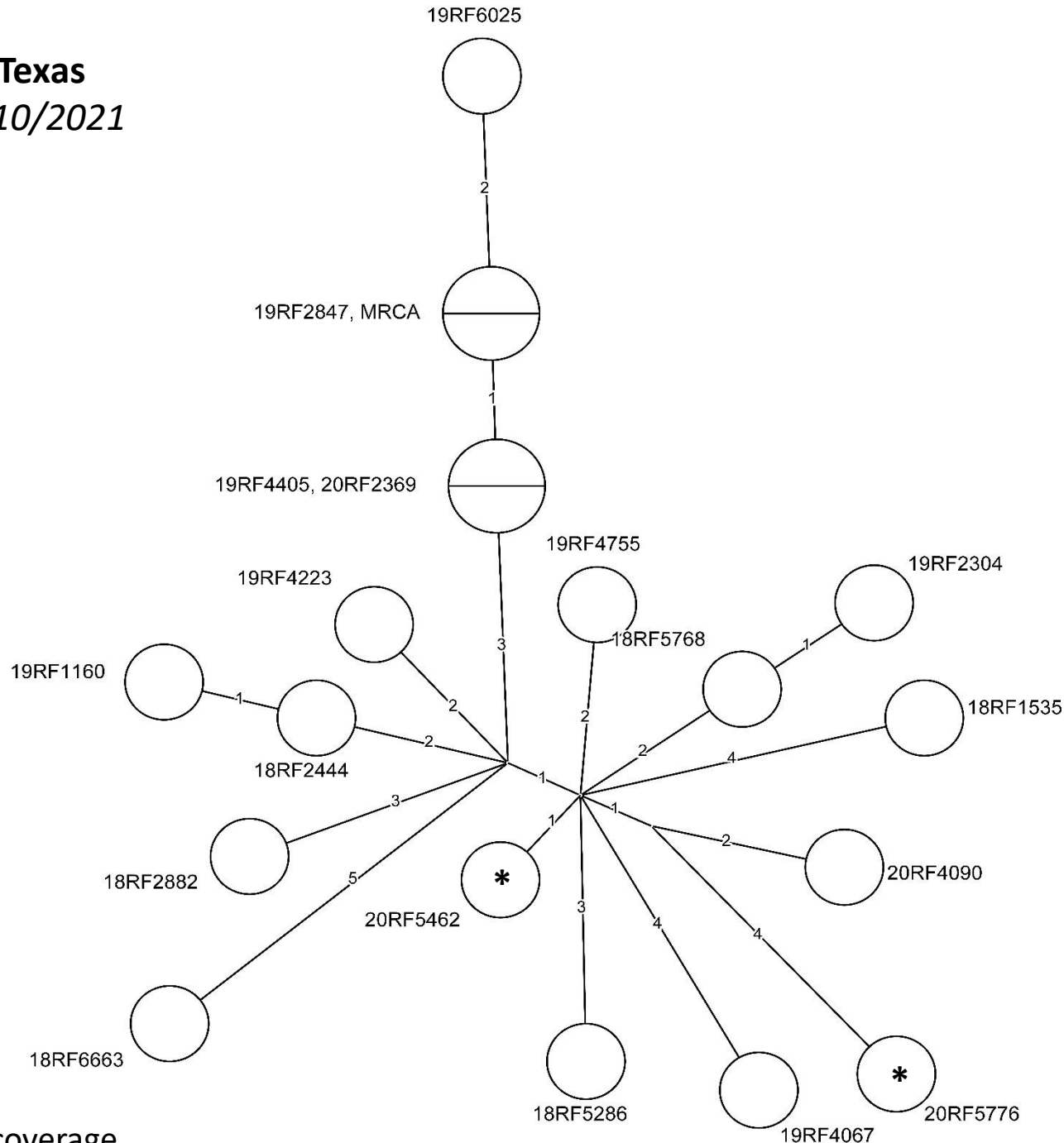
Red text- new targets in WGS not covered by PSQ

	Drug	PSQ loci	Added <b>WGS</b> loci
<b>1<sup>st</sup> line</b>	Isoniazid	<i>katG, fabG1/inhA, ahpC</i>	= PSQ* - <i>ahpC</i>
	Rifampicin	<i>rpoB</i>	= PSQ*
	Pyrazinamide	<i>pncA</i> ( <i>M. bovis</i> ID only)	= PSQ* + <i>pncA</i> (PZA R)
	Ethambutol	-	<i>embA, embB</i>
<b>2<sup>nd</sup> line</b>	Amikacin/Kanamycin	<i>rrs</i>	= PSQ* + <i>eis</i>
	Capreomycin	<i>rrs</i>	= PSQ* + <i>tlyA</i>
	Fluoroquinolones	<i>gyrA</i>	= PSQ* + <i>gyrB</i>
	Ethionamide	<i>fabG1/inhA</i>	= PSQ* + <i>ethA</i>
<b>Other</b>	Bedaquiline	-	<i>Rv0678 (mmpR), pepQ, atpE, mmpL5, mmpS5</i>
	Clofazimine	-	<i>Rv0678 (mmpR), pepQ, atpE, mmpL5, mmpS5</i>
	Linezolid	-	<i>rplC, rrl</i>
<b>MTBC ID confirmation</b>		Yes	Yes

\*For targets included in PSQ, the genomic ranges covered in WGS DST assay are broader and include FL genes & promoter regions (with some exceptions for reporting ranges).

**MTBC000025 in Texas**  
*Results received 2/10/2021*

*Analysis updated with  
20RF5776 and 20RF5462  
(isolates from the same patient)*



**Excluded isolates:**

- 19RF3857 – contaminated
- 18RF6976 – contaminated
- 18RF6084 – contaminated
- 18RF4713 – contaminated
- 18RF3636 – low sequence coverage

*\*Isolates denoted with an asterisk are from the same patient. Isolate 20RF5462 was collected from a sputum specimen and 20RF5776 was collected from a urine sample.*

# Additional DST

- Very few laboratories perform conventional testing on the newer antibiotics
- Molecular targets typically relied upon
- Wadsworth will report clinical breakpoint results for bedaquiline (1.0 µg/mL), clofazimine (1.0 µg/mL), and linezolid (1.0 µg/mL) by MGIT 960
- Can also perform MICs



# Thank you!

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